

The Quorum-Quenching Metallo- γ -lactonase from *Bacillus thuringiensis* Exhibits a Leaving Group Thio Effect[†]

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ABSTRACT: Lactone-hydrolyzing enzymes derived from some *Bacillus* species are capable of disrupting quorum sensing in bacteria that use *N*-acyl-L-homoserine lactones (AHLs) as intercellular signaling molecules. Despite the promise of these quorum-quenching enzymes as therapeutic and anti-biofouling agents, the ring opening mechanism and the role of metal ions in catalysis have not been elucidated. Labeling studies using ¹⁸O, ²H, and the AHL lactonase from *Bacillus thuringiensis* implicate an addition–elimination pathway for ring opening in which a solvent-derived oxygen is incorporated into the product carboxylate, identifying the alcohol as the leaving group. ¹H NMR is used to show that metal binding is required to maintain proper folding. A thio effect is measured for hydrolysis of *N*-hexanoyl-L-homoserine lactone and the corresponding thiolactone by AHL lactonase disubstituted with alternative metal ions, including Mn²⁺, Co²⁺, Zn²⁺, and Cd²⁺. The magnitude of the thio effect on *k*_{cat} values and the thiophilicity of the metal ion substitutions vary in parallel and are consistent with a kinetically significant interaction between the leaving group and the active site metal center during turnover. X-ray absorption spectroscopy confirms that dicobalt substitution does not result in large structural perturbations at the active site. Finally, substitution of the dinuclear metal site with Cd²⁺ results in a greatly enhanced catalyst that can hydrolyze AHLs 1600–24000-fold faster than other reported quorum-quenching enzymes.

Chronic lung infections are a major health concern for patients with cystic fibrosis (1). *Pseudomonas aeruginosa* and *Burkholderia cepacia* are two of the most common bacterial pathogens that can form recalcitrant mixed species biofilms on lung surfaces (2, 3). The *N*-acyl-L-homoserine lactone (AHL)-mediated quorum-sensing pathways of these bacteria have been experimentally linked to their coordinated expression of virulence factors and to mature biofilm development (4). To combat these infections, “quorum-quenching” reagents that can disrupt the quorum-sensing pathways of these organisms have been suggested as novel therapeutics (5). In addition to small molecule antagonists (6), several classes of enzymes capable of catalytically metabolizing AHLs have been reported. Enzymes that cleave

the *N*-acyl bond of AHLs (AHL acylases, Scheme 1a) have been found in several different types of bacteria, including *Ralstonia* strain XJ12B, pseudomonads, and a *Streptomyces* species (7–9). Alternatively, humans and other mammals carry several paraoxonase isozymes capable of breaking a different bond by catalyzing a hydrolytic ring opening (Scheme 1b) (10, 11). However, AHLs are not the preferred substrates of human paraoxonases, which have a 10²–10⁶-fold higher specific activity for non-AHL substrates (11).

The most active quorum-quenching agents discovered to date are naturally occurring AHL lactonases (metallo- γ -lactonases) from certain *Bacillus* species, which can block quorum-sensing-dependent processes in several in vivo systems by catalyzing the hydrolytic ring opening of AHLs (12–14). For example, expression of AHL lactonase in the human pathogen *P. aeruginosa* resulted in large decreases in levels of virulence gene expression and swarming motility (15). Expression of AHL lactonase in *Burkholderia* species can also attenuate virulence (16, 17). In agricultural applica-

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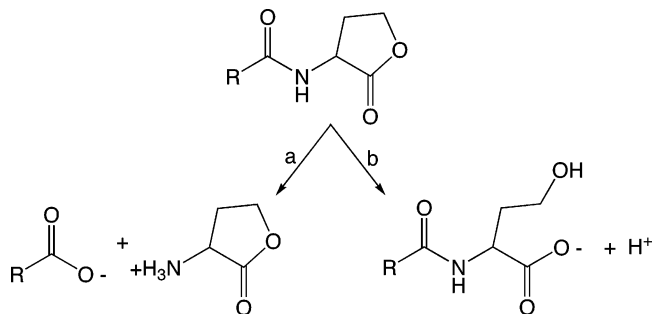
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¹ Abbreviations: AHL, *N*-acyl-L-homoserine lactone; HSL, homoserine lactone; Hcy, homocysteine; Hse, homoserine; MBP, maltose binding protein; LB, Luria-Bertani; PCR, polymerase chain reaction; HCTL, homocysteine thiolactone; SDS, sodium dodecyl sulfate; PAGE, polyacrylamide gel electrophoresis; OD₆₀₀, optical density at 600 nm; ICP-MS, inductively coupled plasma mass spectrometry; *aiiA*, gene for AHL lactonase; TEV, tobacco etch virus; TB, Terrific Broth; MWCO, molecular weight cutoff; HIC, hydrophobic interaction chromatography; 1D, one-dimensional; ESI-HRMS, electrospray ionization high-resolution mass spectrometry; TLC, thin layer chromatography; EXAFS, extended X-ray absorption fine structure; NMR, nuclear magnetic resonance.

Scheme 1: Enzymic Hydrolysis of Quorum-Sensing AHL Signals by (a) AHL Acylases, (b) Paroxonases and AHL Lactonases



tions, the expression of AHL lactonase in transgenic tobacco leaves and potatoes significantly enhances resistance against *Erwinia carotovora* infections, a pathogen reliant on AHL-mediated quorum sensing for its virulence (13). The monomeric AHL lactonase produced by *Bacillus thuringiensis* belongs to the metallo- β -lactamase superfamily and is purified with two tightly bound metal ions shown to be essential for activity (18, 19).

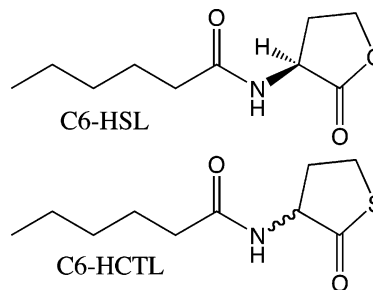
Despite the promise of this enzyme as a potential therapeutic agent, and the publication of several high-resolution X-ray crystal structures (18, 20), little is understood about the AHL lactonase mechanism. The role of metal ions in the catalytic mechanism has been disputed (21), and the mechanism that this enzyme uses to open the AHL ring is undefined. A larger question of significant interest to the entire metallo- β -lactamase superfamily is whether the dinuclear metal sites in these enzymes can use their metal ions for leaving group stabilization (22). The studies described herein provide evidence consistent with an addition–elimination pathway for ring opening, identify the product's alcohol as the leaving group, show that the metal ions associated with AHL lactonase are required for proper protein folding, and use a thio effect to reveal a kinetically significant interaction between the leaving group and the metal center during turnover. These studies establish a foundation for designing more effective therapeutic and anti-biofouling biocatalysts for use against quorum-sensing organisms.

MATERIALS AND METHODS

Unless otherwise noted, all chemicals were obtained from Sigma-Aldrich Chemical Co. (St. Louis, MO) and all restriction enzymes were acquired from New England BioLabs (Beverly, MA). D₂O (99.9%) was purchased from Cambridge Isotope Laboratories and H₂¹⁸O (95%) from Isotec (Miamisburg, OH). Metal analysis of purified proteins was completed using inductively coupled plasma mass spectrometry as described previously (19). The catalytic activities of dizinc and alternative metal-containing AHL lactonases were quantified using a continuous spectrophotometric phenol red-based assay as described previously (19).

Enzyme-Catalyzed Incorporation of ¹⁸O from H₂¹⁸O into the *N*-Hexanoyl-*L*-homoserine Product. A reaction buffer (1 mL) containing 50% H₂¹⁸O and 50% H₂¹⁶O was prepared by mixing KH₂PO₄ buffer (40 mM, 0.47 mL) in H₂¹⁶O (pH 7.4) with H₂¹⁸O (95%, 0.53 mL). To this solution was added *N*-hexanoyl-(*S*)-homoserine lactone [C6-HSL, synthesized as reported previously (19)] (0.5 M, 20 μ L) in a methanol stock

solution. AHL lactonase (6.5 μ g) was added, and the mixture was allowed to react at room temperature for 1 h after which the reaction was stopped by freezing the sample in liquid N₂ and the solvent removed under vacuum using a speedvac (Eppendorf). The remaining solid residue was redissolved in D₂O (99.9%, 1 mL) and incubated at 25 °C for 1 day before being analyzed by ¹³C NMR spectroscopy to determine the resulting peaks for product: ¹³C NMR (D₂O, 500 MHz) δ 13.39, 21.86, 25.21, 30.71, 35.93, 58.84, 62.70, 72.27, 177.05, 179.11, 179.13.



¹H NMR Spectroscopy of Reaction Products after Hydrolysis in D₂O. Products resulting from enzymic and nonenzymic hydrolysis of C6-HSL in 99.9% D₂O (Cambridge Isotope Laboratories) and KH₂PO₄ buffer (20 mM, pH 7.86) were compared. The pH was measured using a standard electrode and corrected using the equation pD = pH + 0.4 (23). Substrate was added from a stock solution of C6-HSL (0.5 M in methanol-*d*₄). Nonenzymic alkaline hydrolysis of C6-HSL (83.3 mM) was carried out in the buffer described above upon addition of NaOH (105 mM, 1.25 equiv). ¹H NMR spectroscopy was performed using a Varian 300 MHz spectrometer before and after hydrolysis. A comparison of the peak integration values for the terminal methyl group in the C6 alkyl chain (3H, 0.9 ppm) and the lactone ring's α proton (1H, 4.3 and 4.1 ppm for the closed and open lactone, respectively) was used to determine whether exchange between hydrogen and solvent deuterium occurs at the α position. Investigation of the enzymic reaction utilized C6-HSL (20 mM) in the same deuterated buffer solution described above. ¹H NMR spectroscopy was carried out on the sample prior to addition of the enzyme. After addition of dizinc AHL lactonase (0.48 mg/mL), the solution was incubated (25 °C), and ¹H NMR spectra were obtained at various time points (1, 2, and 20 h). Comparison of the peak integration values for the terminal methyl group in the C6 alkyl chain and for the lactone α proton was completed as described above.

¹H NMR Spectroscopy of Dinuclear Zinc and Apo AHL Lactonase. Dinuclear zinc AHL lactonase (final concentration of 1.6 mg/mL) was prepared as described previously (19) and dialyzed (12–14 kDa MWCO, Spectrapor) against Chelex (Bio-Rad)-treated KH₂PO₄ buffer (20 mM, pH 7.4), with addition of D₂O (10%, v/v). Samples of apo AHL lactonase (final concentration of 1.6 mg/mL) were prepared as described previously (19) and exchanged into the same buffer described above. One-dimensional (1D) ¹H NMR spectra of the dizinc and apo proteins were recorded using a 500 MHz Varian Inova NMR spectrometer equipped with a triple-resonance probe and *z*-axis pulsed field gradient. In each case, the NMR signal from the solvent was suppressed by presaturation.

Synthesis of *N*-Hexanoylhomocysteine Thiolactone (C6-HCTL). In a procedure similar to that reported elsewhere (24), triethylamine (3.0 mL, 21 mmol) was added to a stirred suspension of racemic DL-homocysteine thiolactone hydrochloride (3.0 g, 20 mmol) in dimethylformamide (350 mL) at 0 °C. Hexanoyl chloride (3.2 mL, 23 mmol) was added dropwise, and the mixture was allowed to react at room temperature with stirring for 1.5 h. Solvent was removed by rotary evaporation with heating at ≤ 50 °C. The residue was dissolved in 2-propanol and purified by column chromatography on silica gel using 2-propanol as the mobile phase. The solvent was removed by rotary evaporation at 20 °C. The remaining residue was dissolved in CH_2Cl_2 and washed sequentially with saturated NaHCO_3 , KHSO_4 (1 M), and saturated NaCl. The organic layer was dried over anhydrous MgSO_4 , followed by removal of solvent by rotary evaporation to leave the final product, which shows as one spot by TLC: $R_f = 0.83$ in 2-propanol; $T_m = 50\text{--}53$ °C (uncorrected); ^1H NMR (300 MHz, $\text{DMSO}-d_6$) δ 0.86 (t, 3H), 1.25 (m, 4H), 1.48 (m, 2H), 2.04–2.12 (m, 3H), 2.36 (m, 1H), 3.25 (m, 1H), 3.35 (m, 1H), 4.57 (m, 1H), 8.12 (broad, 1H); ^{13}C NMR (75 MHz, $\text{DMSO}-d_6$) δ 13.86, 21.86, 24.85, 26.68, 30.18, 30.86, 35.17, 58.04, 172.30, 205.50; EI-HRMS $\text{MH}^+_{\text{calc}} = 216.0980$, $\text{MH}^+_{\text{obs}} = 216.1060$.

Expression and Purification of Metal-Substituted AHL Lactonases. To incorporate different metal ions into AHL lactonase, expression conditions were altered from previous procedures (19). Briefly, *Escherichia coli* BL21(DE3) cells harboring the pMAL-t-ahA expression plasmid (19) were incubated at 37 °C with shaking in M9 minimal medium [Na_2HPO_4 (47 mM), KH_2PO_4 (22 mM), NaCl (8.5 mM), NH_4Cl (18.7 mM), MgSO_4 (2.0 mM), CaCl_2 (0.1 mM), and glucose (0.4%, w/v)] supplemented with ampicillin (100 $\mu\text{g}/\text{mL}$) until cells reached an OD_{600} of 0.5–0.7. Cells were harvested by centrifugation and then resuspended in fresh M9 minimal medium containing IPTG (0.3 mM), ampicillin (100 $\mu\text{g}/\text{mL}$), and one of the following metal salts (0.5 mM): ZnSO_4 , CoCl_2 , CdCl_2 , or MnSO_4 . Induction at 25 °C was continued with shaking for an additional 16 h, followed by harvesting by centrifugation at 8275g and 4 °C for 7 min. Cell pellets from 2 L of expression culture were resuspended in 50 mL of Tris-HCl buffer (20 mM, pH 7.4) with NaCl (100 mM) and stored at -20 °C until they were used.

Unless otherwise noted, all purification procedures were carried out at 4 °C. BioLogic LP protein purification systems (Bio-Rad, Hercules, CA) were used for all chromatographic procedures. Thawed cell pellets were resuspended in 200 mL of Tris-HCl buffer (10 mM, pH 7.4) with NaCl (100 mM). Cells were lysed by sonication, and the cell debris was removed as described previously (19). The resulting supernatant was loaded onto a DEAE-Sepharose FF column (2.5 cm \times 20 cm) equilibrated with buffer A [20 mM Tris-HCl and 5 mM NaCl (pH 7.4)], and the column was washed with 200 mL of buffer A made to 160 mM NaCl. The salt concentration was subsequently increased to 250 mM (NaCl) using a linear gradient. The fractions containing active fusion protein generally eluted at a conductivity near 20 mS/cm (~ 200 mM NaCl). Pooled fractions containing maltose-binding protein (MBP)–AHL lactonase fusion protein were split into two portions and loaded in parallel on two columns (2.5 cm \times 10 cm) of amylose–agarose resin (New England BioLabs) at a flow rate of 0.7 mL/min. Each column was

washed with 2–3 column volumes of buffer A made to 100 mM NaCl at a flow rate of <1 mL/min, and the fusion protein was subsequently eluted at a flow rate of 1.5 mL/min using buffer A made to 100 mM NaCl and 10 mM maltose.

The MBP–AHL lactonase fusion proteins prepared with ZnSO_4 or CoCl_2 were subsequently cleaved with tobacco etch virus protease, and the untagged enzyme was purified as described previously (19). During preliminary cleavage experiments, the MBP–AHL lactonase fusion proteins prepared using CdCl_2 or MnSO_4 were observed to gradually bind zinc, present in buffers only at trace concentrations, at the expense of either cadmium or manganese (data not shown). To avoid the resulting metal heterogeneity in these purified proteins, the lactonases that were grown in media supplemented with either CdCl_2 or MnSO_4 were not cleaved but rather used as the MBP fusion proteins for the remaining experiments. It has previously been demonstrated that AHL lactonase is monomeric (18) and that the presence of the MBP tag does not significantly affect catalysis (19).

EXAFS of Dinuclear Cobalt AHL Lactonase. Samples of dicobalt AHL lactonase (ca. 1 mM) containing 10% (v/v) glycerol were loaded in Lucite cuvettes with 6 μm polypropylene windows and frozen rapidly in liquid nitrogen. X-ray absorption spectra were measured at the National Synchrotron Light Source (NSLS), beamline X9B, with a Si(111) double-crystal monochromator; harmonic rejection was accomplished using a Ni focusing mirror. EXAFS data collection and reduction were performed according to published procedures (19, 25). Spectra were collected on two independently prepared samples. As each data set gave similar results, the two data sets were averaged. The spectra shown (see below) represent the average of the two data sets (16 total scans, 13 detector channels per scan). Background-subtracted EXAFS spectra were Fourier transformed over the k range of $1\text{--}13.6$ \AA^{-1} ; Fourier-filtered EXAFS data ($\Delta R = 0.8\text{--}2.0$ \AA for first-shell fits, $\Delta R = 0.1\text{--}4.5$ \AA for multiple scattering fits) were then fit using amplitude and phase functions, $A_s(k) \exp(-2R_{\text{as}}/\lambda)$ and $\phi_{\text{as}}(k)$, respectively, that were calculated using FEFF version 8.00 (26). The Co–N scale factor, S_c , and the threshold energy, ΔE_0 , were determined by fitting the experimental spectrum for bis-trispyrazolylborato cobalt(II) (25). These two parameters were then held fixed at their optimal values ($S_c = 0.78$ and $\Delta E_0 = -21$ eV) in subsequent fits to protein data, which led to variation of only R_{as} and σ_{as}^2 for a given shell. Metal–metal (Co–Co) scattering was modeled by fitting calculated amplitude and phase functions to the experimental EXAFS of $\text{Co}_2(\text{sapln})_2$.

RESULTS

Enzyme-Catalyzed Incorporation of ^{18}O from H_2^{18}O into the *N*-Hexanoyl-*L*-homoserine Product. AHL lactonase was used to hydrolyze C6-HSL in a mixture of 50% H_2^{18}O and 50% H_2^{16}O to detect an isotope-induced shift in the ^{13}C resonance for the carbon adjacent to the site of incorporation (27). To rule out nonenzymic exchange of H_2^{18}O into the product, after a short incubation (1 h) of substrate with the enzyme, the samples were dried and resuspended in D_2O followed by a long incubation (1 day) before analysis. The presence of a characteristic ^{18}O isotope-induced ^{13}C shift (see

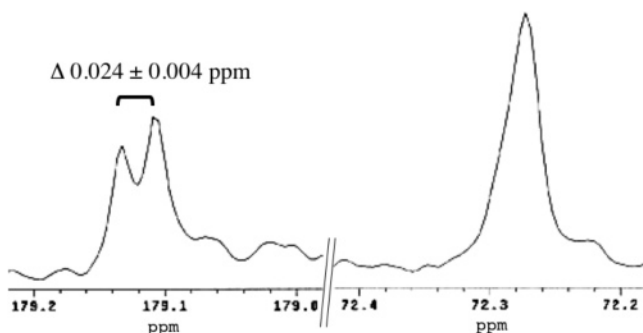


FIGURE 1: Partial ^{13}C NMR spectra of the ^{18}O -labeled product. C6-HSL was hydrolyzed by AHL lactonase in 50% H_2^{18}O and 50% H_2^{16}O , and two selected regions of the ^{13}C NMR spectrum of the resulting product, *N*-hexanoyl-L-homoserine, are shown, corresponding to the carboxylic carbon (~ 179 ppm) and the C_γ methylene carbon (~ 72 ppm).

below) indicates that there was no significant washout of this position during the D_2O incubation following enzymic cleavage. This result is consistent with reports that oxygen exchange reactions with acetic acid in water are acid-catalyzed, and only very slow exchange occurs when pH values are greater than 6.0 (28). For the ring-opened product obtained after incubation with AHL lactonase, the ^{13}C resonance peak for the C_γ methylene carbon at 72.272 ppm and a split peak for the ring carbonyl carbon at 179.108 and 179.132 ppm are shown in Figure 1. The magnitude of the new upfield isotope shift observed at 179 ppm is 0.024 ± 0.004 ppm relative to the resonance position of the unlabeled ^{16}O -containing product, consistent with incorporation of one ^{18}O atom into the product carboxylate (27, 29).

^1H NMR Spectroscopy of Reaction Products after Hydrolysis in D_2O . To determine whether the lactone α proton is exchanged with solvent deuterium during catalysis, non-enzymic and AHL lactonase-catalyzed hydrolysis of C6-HSL were completed in deuterated buffer and the reaction products analyzed by ^1H NMR spectroscopy. In both cases, peak integration values were compared between the protons of the terminal methyl group of the alkyl chain and the α proton (see the Discussion). These peaks are well-resolved, and the protons of the methyl group should not readily exchange with solvent, allowing these values to be set equivalent to three protons. In the nonenzymic control reaction, before hydrolysis, the methyl protons (3.0H, 0.73 ppm) integrated with a 3:1 ratio with the α proton (1.1H, 4.25 ppm) of the substrate. After addition of hydroxide, characteristic chemical shifts indicated complete ring opening (data not shown), and the methyl protons (3.0H, 0.71 ppm) integrated with approximately the same ratio with the α proton of the product (0.8H, 4.06 ppm). In the enzyme-catalyzed reaction, complete hydrolysis was observed at the first time point (1 h) and the methyl protons (3.0H, 0.72 ppm) integrated with a 3:1 ratio with the α proton of the product (0.9H, 4.08 ppm). To ensure no further reaction, the enzymic reaction was analyzed at later time points (2 and 20 h). The methyl protons (2 h, 3.0H, 0.72 ppm; 20 h, 3.0H, 0.73 ppm) and the product's α proton (2 h, 1.1H, 4.08 ppm; 20 h, 1.2H, 4.10 ppm) remained in a 3:1 ratio, indicating that there is not significant exchange of the α proton with solvent deuterium during turnover.

^1H NMR Spectroscopy of Dinuclear Zinc AHL Lactonase and Apo AHL Lactonase. To determine whether removal of

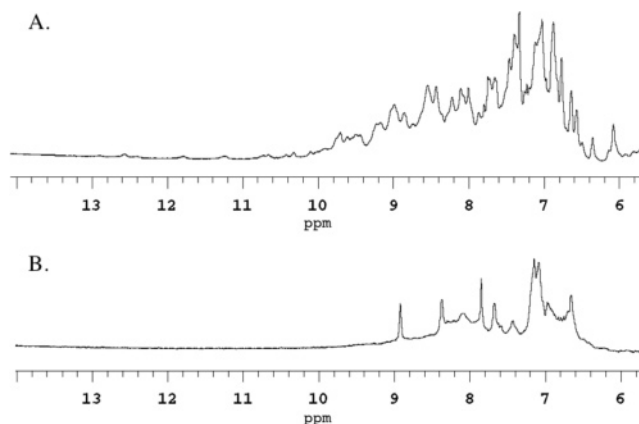


FIGURE 2: Partial one-dimensional ^1H NMR spectra of dinuclear zinc AHL lactonase (A) and apo AHL lactonase (B), indicating significant unfolding after removal of metal ions.

Table 1: Metal Content of Purified Monomeric AHL Lactonase Grown in Minimal Medium with Different Metal Supplements

metal supplement	manganese (equiv)	cobalt (equiv)	zinc (equiv)	cadmium (equiv)
MnSO_4	1.8	<0.01	0.1	0.06
CoCl_2	<0.01	1.9	0.2	<0.01
ZnSO_4	<0.01	<0.01	2.0	<0.01
CdCl_2	<0.01	<0.01	0.04	2.0

metal ions from AHL lactonase resulted in any protein unfolding, 1D ^1H NMR spectra were compared between dizinc and apo AHL lactonase for differences in downfield-shifted resonances typical of folded proteins (30, 31). 1D ^1H NMR spectroscopy is commonly used to qualitatively determine whether a protein is folded or unstructured (30). Dizinc AHL lactonase appears to be well-folded, exhibiting peaks for backbone amides shifted downfield by 8.5 ppm (Figure 2A). After removal of the metal ions, most of these downfield peaks disappeared and broader features were present near 8.3 ppm, the characteristic shift of backbone amides in random coils (Figure 2B). It is unlikely that the chelator itself causes unfolding because only modest concentrations are used (2 mM) and because re-addition of metal ions (19), but not removal of the chelators (this work), can restore enzyme activity. These observations are consistent with a significant structural destabilization of AHL lactonase upon, or after, removal of the active site metal ions.

Expression and Purification of Metal-Substituted AHL Lactonases. Yields of purified untagged AHL lactonase (29 kDa) from 2 L expression cultures were typically greater when the induction medium was supplemented with CoCl_2 (40 mg) than with ZnSO_4 (7 mg). Yields of purified MBP-AHL lactonase fusion proteins (73 kDa) from 2 L expression cultures were lower when the induction medium was supplemented with CdCl_2 (10 mg) or MnSO_4 (22 mg). Metal analysis was performed as previously described (19) and confirmed that enzymes grown in each metal supplement resulted in a purified protein containing approximately 2 equiv of the supplemented metal ion per monomer (Table 1). Several independent expression and purification trials confirmed the reproducibility of these procedures for preparing AHL lactonase that is homogeneously substituted with alternative divalent metal ions.

Steady-State Kinetics of Metal-Substituted AHL Lactonases. Using a previously reported (19) phenol red-based

Table 2: Steady-State Kinetic Constants for Hydrolysis of Substrates with Oxygen (C6-HSL, O) or Sulfur (C6-HCTL, S) Leaving Groups by AHL Lactonase Disubstituted with Various Divalent Metal Ions^a

metal supplement	C6-HSL		C6-HCTL		$k_{\text{cat}}(\text{O})^b/k_{\text{cat}}(\text{S})$	$K_{\text{M}}(\text{O})/K_{\text{M}}(\text{S})$
	k_{cat} (s ^{−1})	K_{M} (mM)	k_{cat} (s ^{−1})	K_{M} (mM)		
MnSO ₄	330 ± 20	0.18 ± 0.04	500 ± 20	0.39 ± 0.05	0.66 ± 0.07	0.4 ± 0.2
CoCl ₂	510 ± 10	0.36 ± 0.04	198 ± 8	6.7 ± 0.7	2.6 ± 0.2	0.054 ± 0.006
ZnSO ₄	91 ± 3	5.6 ± 0.6	4.1 ± 0.7	36 ± 10	22 ± 5	0.16 ± 0.06
CdCl ₂	480 ± 20	0.24 ± 0.03	5.0 ± 0.5	0.12 ± 0.02	100 ± 20	2.0 ± 0.6

^a Reactions were carried out at pH 7.4 and 28 °C. ^b Ratios compare the relative steady-state kinetic constants for hydrolysis of substrates with an oxygen (O) leaving group, C6-HSL, or with a sulfur (S) leaving group, C6-HCTL.

spectrophotometric activity assay, the steady-state rate constants were measured for hydrolysis of C6-HSL and C6-HCTL at 28 °C and pH 7.4 by AHL lactonase disubstituted with manganese, cobalt, zinc, or cadmium (Table 2). Progress curves remained linear over the time scale used for calculating initial rates, typically ~0.2 min. The maximum substrate concentrations used in these experiments were at least 5-fold higher than K_{M} values, except for hydrolysis of C6-HCTL by dizinc AHL lactonase, where the high K_{M} and the limited aqueous solubility of the thiolactone allowed assays with only a maximum near 40 mM substrate.

In general, rankings of the absolute values for steady-state rate constants do not obviously correlate with the Lewis acid strength or thiophilicity of the substituted metal ions. For the C6-HSL substrate, k_{cat} values increased upon substitution in the following order: $\text{Zn}^{2+} \ll \text{Mn}^{2+} < \text{Cd}^{2+} < \text{Co}^{2+}$. K_{M} values decreased with a different rank order: $\text{Zn}^{2+} \gg \text{Co}^{2+} > \text{Cd}^{2+} > \text{Mn}^{2+}$. These absolute rankings are different for the sulfur-containing C6-HCTL substrate, which exhibited increasing k_{cat} values upon substitution ($\text{Zn}^{2+} < \text{Cd}^{2+} \ll \text{Co}^{2+} < \text{Mn}^{2+}$) and decreasing K_{M} values ($\text{Zn}^{2+} > \text{Co}^{2+} > \text{Mn}^{2+} > \text{Cd}^{2+}$). These results are somewhat surprising because, in all cases, disubstitution with zinc results in the least effective catalyst. In contrast, zinc appears to be the optimum metal ion for catalysis by a related metallo- β -lactamase (32).

While the trends of these absolute values are difficult to interpret, it is more instructive to compare the ratio of rate constants for the oxygen- and sulfur-containing substrates (O:S ratios in Table 2) which can be used to isolate the effects of metal substitution on the leaving group position (see the Discussion). The O:S ratio of k_{cat} values for C6-HSL and C6-HCTL showed a clear trend, varying by 150-fold, with the following rank order: $\text{Mn}^{2+} < \text{Co}^{2+} < \text{Zn}^{2+} < \text{Cd}^{2+}$. Under our experimental conditions, product inhibition did not significantly contribute to the observed thio effect (Supporting Information). This ranking of O:S k_{cat} ratios matched the trend of reported stability constants for these metal ions binding to small molecule thiols [$\text{Mn}^{2+} < \text{Co}^{2+} < \text{Zn}^{2+} < \text{Cd}^{2+}$ (33–36)] and is consistent with an apparent correlation between the ability of these metal ions to bind thiols and their ability to perturb the k_{cat} values of thiolactones as compared to the oxygen congeners.

EXAFS of Dinuclear Cobalt AHL Lactonase. Our previous EXAFS studies of dizinc AHL lactonase revealed a dinuclear structure, with an average of five N/O donors per zinc ion, including 2 ± 0.5 His ligands per zinc ion. The Zn–Zn distance of 3.32 Å was taken as evidence of the presence of two bridging groups, including a carboxylate and a solvent molecule (19). To determine whether the changes in reactivity upon metal substitution are a manifestation of structural

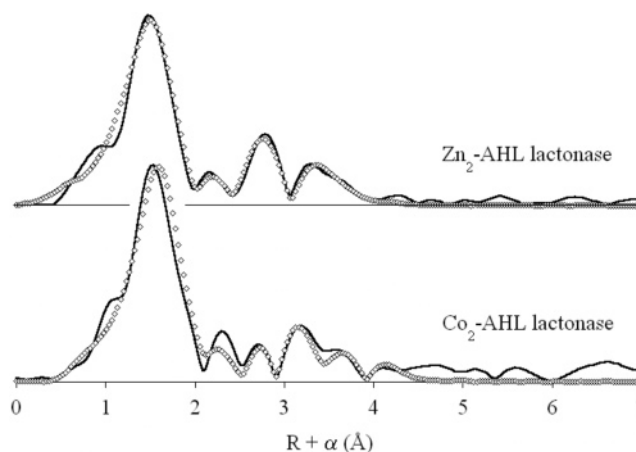


FIGURE 3: Fourier-transformed EXAFS data (—) for dizinc (top) (19) and dicobalt (bottom) AHL lactonase, and corresponding best fits (◇).

perturbations, we examined the EXAFS of the dicobalt enzyme.

The data presented in Figure 3 are consistent with similar structures for the dizinc and dicobalt enzymes (the original dizinc spectrum is included for comparison). First-shell fits to the dicobalt data, summarized in Table 3, indicate an average of 5 ± 1 N/O donors per cobalt ion, at a distance of 2.03 Å. These results are indistinguishable from those observed for the dizinc enzyme, indicating identical coordination numbers for the Zn and Co ions. Attempts to fit the first shell with a mixture of nitrogen and oxygen donors did not lead to significant improvement in the fit residuals.

Multiple scattering fits indicate the presence of an average of 2 ± 0.5 His ligands per Co(II) ion, also in accord with the dizinc EXAFS (19). These fits adequately reproduce the position of the four prominent outer shell features but poorly model the intensity pattern, particularly for the feature at $R + \alpha = 3.1$ Å. Inclusion of a Co–Co interaction dramatically improves the appearance of the fit, more closely matching the observed outer shell intensities. More importantly, addition of the Co–Co interaction (an increase in the number of variable parameters in the fit from 10 to 12) leads to a 27% improvement in the fit residual. The Co–Co distance refines to 3.55 Å, which is significantly longer than the 3.32 Å Zn–Zn distance observed for dizinc AHL lactonase. This can be ascribed, in part, to the slightly larger (0.06 Å) covalent radius of Co(II) versus Zn(II), which predicts an increase of up to 0.12 Å in the metal–metal distance. The remaining 0.1 Å increase is at most a minor perturbation of the active site structure.

Table 3: EXAFS Fitting Results for Dicobalt AHL Lactonase^a

fit	scatterer ^b	path ^c	R_{as} (Å)	σ_{as}^2 ^d	R_f ^e	R_u ^e
1	5 N/O		2.03	7.2	27	267
2	5 N/O		2.03	6.0	199	201
	2 His	C1	2.98	9.0		
		C1–N1	3.35	6.0		
		C2–N1	4.22	15.		
		C1–N2–N1	4.67	14.		
3	5 N/O		2.03	6.0	146	157
	2 His	C1	3.02	6.4		
		C1–N1	3.38	16.		
		C2–N1	4.24	6.6		
		C1–N2–N1	4.68	13.		
	1 Co	Co	3.55	9.0		

^a Values of R_{as} and σ_{as}^2 are for fits to filtered data, as described in Materials and Methods. Fits to unfiltered data gave similar results.

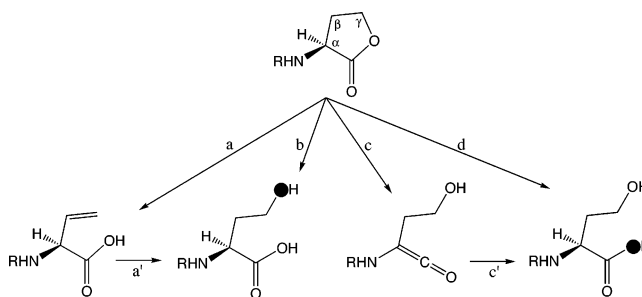
^b Integer coordination number giving the best fit. ^c Multiple scattering paths represent combined paths, with labels that correspond to the path with the largest amplitude (19). ^d Mean-square deviation in absorber–scatterer bond length of 10^{-3} Å². ^e Goodness of fit defined as

$$1000 \times \frac{\sum_{i=1}^N \{[\text{Re}(\chi_{i_{\text{calc}}})]^2 + [\text{Im}(\chi_{i_{\text{calc}}})]^2\}}{\sum_{i=1}^N \{[\text{Re}(\chi_{i_{\text{obs}}})]^2 + [\text{Im}(\chi_{i_{\text{obs}}})]^2\}}$$

where N is the number of data points. R_f corresponds to fits to filtered data, and R_u corresponds to fits to unfiltered data. The apparent similarity of the residuals for filtered and unfiltered fits is an artifact of the Fourier filtering process, which converts real data (unfiltered) into complex data (filtered). This results in an effective doubling of the number of points being fit, leading to apparently higher residuals for fits to filtered data.

DISCUSSION

Although AHL lactonase holds potential as a therapeutic and anti-biofouling agent, its enzymic ring opening mechanism has not yet been characterized. Generally, alkaline ester hydrolysis occurs through an addition–elimination pathway (rather than elimination–addition, or an S_N2 pathway). However, lactones are known to have different reactivity than open chain esters (37). For instance, rings with fewer than eight members force the alkyl and acyl groups of the lactone into a *cis* configuration as opposed to the *trans* configuration of open-chained esters, and these lactones hydrolyze more than 200-fold faster than their open chain analogues (38; reviewed in ref 37). Besides the expected attack of hydroxide at the carbonyl carbon, alternative mechanisms for AHL lactonase should also be considered. For example, enzyme-catalyzed ring opening of type B streptogramin macrolactones has been shown to occur via elimination mechanisms rather than hydrolysis (39). The nonenzymatic hydrolysis of lactones can also occur by different mechanisms. If an ionizable α proton is present, this may allow deprotonation and elimination of the alcohol to form a ketene that would subsequently react with water to give product. While this mechanism has not been observed in nonenzymic lactone hydrolysis, it has been observed with particular open chain esters (37, 40, 41). Even differences in the site of bond cleavage have been noted. Under neutral conditions, four-member lactone rings have been shown to open between the methylene carbon and the ring oxygen rather than between the carbonyl carbon and the ring oxygen (reviewed in ref 41). This unusual cleavage has even been

Scheme 2: Possible AHL Ring-Opening Mechanisms^a

^a Predicted labeling by solvent $H_2^{18}O$ is indicated (●).

observed during nonenzymic acid hydrolysis of *N*-benzoyl-homoserine lactone (42). Because the dinuclear metal center of AHL lactonase provides two strong Lewis acids as well as a preformed hydroxide ion, it is not clear whether an acidic or an alkaline hydrolysis mechanism is most appropriate to use as a model. Therefore, the possibility of several alternative pathways for lactone hydrolysis justifies the investigation of the particular ring opening mechanism catalyzed by AHL lactonase.

In light of the known possibilities, we considered four potential ring opening mechanisms for AHL lactonase (Scheme 2): (a) elimination of the acetate group [the resulting alkene product has not been detected in previous analysis of reaction products (19, 21, 43), but elimination might be followed by hydration (path a') to yield the expected product], (b) attack of hydroxide at the γ carbon with the acetate acting as a leaving group, (c) deprotonation of the α proton and elimination of the alcohol to form a ketene, which could readily react (path c') with solvent to yield product, and (d) hydroxide attack at the carbonyl carbon.

Both pathways a and b are predicted to incorporate solvent oxygen into the product alcohol, but pathways c and d predict incorporation of solvent oxygen into the product's carboxylate. To determine the actual site of incorporation, C6-HSL was hydrolyzed by AHL lactonase in a buffered solution containing 50% $H_2^{18}O$ and 50% $H_2^{16}O$, and the resulting product was analyzed by natural abundance ^{13}C NMR spectroscopy. Incorporation of an ^{18}O isotope adjacent to a carbon atom is known to introduce a small but reproducible change in the adjacent carbon's chemical shift (27). After hydrolysis, the product's γ carbon exhibits only a single symmetrical peak (Figure 1), with no indication of a neighboring ^{18}O label. However, the product's carbonyl carbon peak is split into two, with approximately 50% of the sample shifted upfield by 0.024 ± 0.004 ppm. The magnitude of the ^{18}O -induced isotope shift agrees with that observed previously with carboxylic acids and indicates that only one ^{18}O atom is incorporated into the product (29). The observed site of incorporation of solvent oxygen into the product carboxylate rather than into the alcohol rules out pathways a and b.

Pathways c and d can be distinguished by washout of the α proton during hydrolysis. Pathway c predicts that the α proton should readily exchange with solvent, but pathway d predicts this proton would be retained. To determine whether the α proton is washed out during turnover, C6-HSL was hydrolyzed by AHL lactonase in buffer containing >95% D_2O and the resulting product analyzed by 1H NMR spectroscopy. The product's α proton integrates well with

the three protons of the terminal methyl group of the alkyl chain and exhibits a 1:3 stoichiometry, indicating that there is no significant exchange at this position during catalysis. This finding disfavors pathway c. These results do not rule out the formal possibility that a general base, not in rapid equilibrium with the solvent, catalyzes both removal of the α proton and reprotonation of the ketene. However, the high pK_a of the α proton and the solvent accessibility of the active site (18) make this possibility unlikely. In sum, isotope labeling patterns are most consistent with an addition–elimination mechanism with attack of a solvent-derived hydroxide at the carbonyl carbon and elimination of an alcohol leaving group.

Another significant unknown in the AHL lactonase mechanism is the role of metal ions. The absolute requirement of metal ions for the catalytic activity of AHL lactonase has been debated (21). Our group [and others (20)] found 2 equiv of Zn^{2+} that copurified with this enzyme and demonstrated that the presence of these metal ions is required for catalytic activity (19). However, loss of activity upon removal of metal ions (a) could be caused by protein unfolding, indicating a structural role for the metal ions, (b) could be caused by removal of these Lewis acids without significant unfolding, suggesting a catalytic role for the metal ions, or (c) could be a combination of these two effects. To determine whether metal ion binding is important for stabilization of the protein fold, we compared qualitative differences between the NMR spectra of dinuclear zinc AHL lactonase and apo AHL lactonase, which no longer contains any bound metal ions (Figure 2). Comparison of 1D 1H NMR spectra is commonly used to qualitatively detect gross changes in protein folding (30). The spectrum of dinuclear zinc AHL lactonase contains many defined peaks, even occurring well above 8.5 ppm, distinctive of the backbone amide protons in folded proteins. In contrast, the spectrum of apo AHL lactonase shows fewer, broader, and less resolved peaks, mostly occurring near 8.3 ppm, indicating a loss (44) of structural integrity upon removal of metal ions. This result is consistent with the dinuclear zinc site serving a structural role in stabilizing the protein fold of AHL lactonase. Indeed, crystal structures of this enzyme show that the dizinc site helps bridge two pseudosymmetrical halves of the overall $\alpha\beta\beta\alpha$ fold (18, 20), and reconstitution experiments with apoprotein and metal ions are slow to reach equilibrium and produce only low yields (19). Other superfamily members have also been shown to rely on metal ion binding for proper folding (44, 45). While these results support a structural role for the bound metal ions, they do not provide any information about whether these metals might also play a role in catalysis. To determine whether the metal ions of AHL lactonase directly participate in catalysis, experiments to track specific metal–substrate interactions that occur throughout the reaction coordinate are underway.

The importance of metal ions in stabilizing the proper fold of AHL lactonase and the identity of the leaving group have now been established. One of the remaining questions is whether the metal center participates in the catalytic mechanism. Most metallo- β -lactamase superfamily members rely on active site metal ions for catalysis (46). One structure of AHL lactonase complexed with homoserine lactone indicates ligand–metal contacts (20), but this likely represents a nonproductive complex. In contrast, early studies of AHL

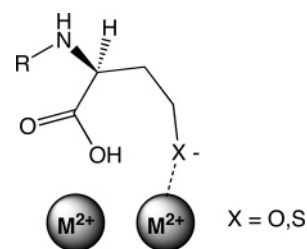


FIGURE 4: Proposed metal–leaving group interaction during lactone and thiolactone hydrolysis by AHL lactonase.

lactonase suggest that metals may not be essential (21), and recently published structures of a related dinuclear zinc alkylsulfatase suggest that the active site metal ions may not interact directly with the substrate (47). Herein, we measured the kinetic thio effect on rates of hydrolysis of lactone and thiolactone substrates by AHL lactonase substituted with metals of varying thiophilicity. If the leaving group does not interact with the metal center, then there should be no change in the ratio of hydrolysis rates for the lactone and thiolactone substrates (O:S ratio) upon metal substitution. However, if the leaving group forms a kinetically significant interaction with the metal center, then it would be reasonable to expect that the ratio of hydrolysis rates should vary in parallel with the thiophilicity of the active site metals. Similar experiments are common in the study of metal-mediated phosphoester hydrolysis (48), and the use of thionolactams (49), thioureas (50), and thionopeptides (34, 51) has also been used to assay for a thio effect on various metallohydrolases. It should be noted that the studies described here are somewhat different from the previous studies because the sulfur-for-oxygen substitution is made at the leaving group position rather than at the carbonyl group.

Steady-state kinetic constants for hydrolysis of C6-HSL and C6-HCTL were determined as catalyzed by AHL lactonase containing 2 equiv each of four different metal ions (Tables 1 and 2): Mn^{2+} , Co^{2+} , Zn^{2+} , and Cd^{2+} . The O:S ratios of K_M values do not show a clear correlation with the ranking of the thiophilicities of these metals, nor is a correlation necessarily expected. However, the O:S ratios of k_{cat} values, which vary by 150-fold, show a strong correlation with the known thiophilicity of the active site metal ions. The rank order of O:S k_{cat} ratios for metal-substituted AHL lactonase ($Cd^{2+} > Zn^{2+} > Co^{2+} > Mn^{2+}$) correlates well with the ranking (52) of the thiophilicities of these metals: $Cd^{2+} > Zn^{2+} > Co^{2+} > Mn^{2+}$ (Table 2). In fact, under saturating conditions, the Mn^{2+} -substituted AHL lactonase, expected to be the least thiophilic species, hydrolyzed the thiolactone faster than the oxygen-containing congener. In contrast, Cd^{2+} -substituted AHL lactonase, expected to be the most thiophilic species, hydrolyzed the oxygen-containing lactone faster than the thiolactone. The observation that the most thiophilic metal displays the slowest rate for thiolactone hydrolysis suggests that breaking the leaving group–metal bond (Figure 4) may be at least partially rate-limiting during steady-state turnover. A similar rate-limiting metal–leaving group interaction has been characterized in related metallo- β -lactamase enzymes (22), although this interaction may be substrate-dependent (53). These findings are consistent with the presence of a kinetically significant interaction between the substrate's leaving group and the active site metal ions of AHL lactonase during turnover. Future kinetic and

spectroscopic studies will address the leaving group–metal interaction of AHL lactonase in greater detail and determine which of the two metal ions is responsible for this interaction.

To ensure that the different metal ion substitutions do not cause large structural changes in the dinuclear site, X-ray absorption spectroscopy of the dinuclear cobalt AHL lactonase (Table 3 and Figure 3) was compared with that reported previously for the dizinc enzyme (19). Fits to the EXAFS data indicate identical coordination spheres for both Co(II) and Zn(II) in the AHL lactonase active site, including an average of five nitrogen/oxygen ligands per metal ion, 2 ± 0.5 of which are histidine residues. The 3.55 Å cobalt–cobalt distance determined for dicobalt AHL lactonase is slightly longer than the 3.32 Å zinc–zinc distance in the dizinc enzyme but is still consistent with formation of a dinuclear cobalt metal center. In sum, no major structural differences in the metal binding site are observed by EXAFS between the dicobalt and dizinc AHL lactonase. Therefore, the observed differences in reactivity between dizinc and dicobalt AHL lactonase are likely due to intrinsic differences in each metal's properties rather than changes in the protein structure.

One unexpected benefit of this study is the finding that different metal ions can greatly enhance catalytic rates, with the Cd^{2+} -substituted enzyme exhibiting a 5-fold increase in k_{cat} , a 23-fold decrease in K_{M} , and a corresponding 125-fold increase in $k_{\text{cat}}/K_{\text{M}}$ for hydrolysis of a naturally occurring substrate, C6-HSL, as compared with the value for the dizinc enzyme. This dicadmium catalyst is orders of magnitude faster than other reported quorum-quenching enzymes. For example, the AHL-acylase AhlM (9) is 3200-fold slower, and human paraoxonases-1, -2, and -3 are 24000-, 1600-, and 9600-fold slower, respectively (11), than dicadmium AHL lactonase.² Further studies will be required to determine whether more physiologically relevant metal ions, such as Fe^{2+} , also afford rate enhancements and whether the non-physiological alternative metals which display greater activity can remain stably incorporated into AHL lactonase when used in a biological milieu.

In conclusion, we have demonstrated that the dinuclear metal site of AHL lactonase is essential both for structural stabilization and for catalysis. Substrate AHL ring opening is shown to be consistent with a proposed addition–elimination mechanism where solvent-derived hydroxide, presumably the hydroxide that bridges the dinuclear zinc site, attacks the lactone's carbonyl and an alcohol leaving group is eliminated. An observed thio effect indicates that a kinetically significant interaction between the substrate's leaving group and the metal center occurs during catalysis and is consistent with proposing a role of the metal center in stabilizing the leaving group, thereby facilitating bond cleavage. Because alternative metal incorporation results in a hyperactive AHL lactonase, this raises the possibility that through an understanding of the structure and mechanism of this enzyme family, more effective quorum-quenching agents can be developed for use as antibacterial and anti-biofouling agents.

² Kinetic characterizations of quorum-quenching lactonases are scant, but the k_{cat} values of dicadmium AHL lactonase are compared with those calculated from the highest reported specific activities of AhlM and PON1–3, assuming saturating optimal AHL substrates and one active site per monomer.

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SUPPORTING INFORMATION AVAILABLE

Product synthesis, product inhibition constants, and comparison of product inhibition with the observed thio effect. This material is available free of charge via the Internet at <http://pubs.acs.org>.

REFERENCES

- Singh, P. K., Schaefer, A. L., Parsek, M. R., Moninger, T. O., Welsh, M. J., and Greenberg, E. P. (2000) Quorum-sensing signals indicate that cystic fibrosis lungs are infected with bacterial biofilms, *Nature* 407, 762–74.
- Smith, R. S., and Iglewski, B. H. (2003) *P. aeruginosa* quorum-sensing systems and virulence, *Curr. Opin. Microbiol.* 6, 56–60.
- Lewenza, S., Visser, M. B., and Sokol, P. A. (2002) Interspecies communication between *Burkholderia cepacia* and *Pseudomonas aeruginosa*, *Can. J. Microbiol.* 48, 707–16.
- Dong, Y. H., and Zhang, L. H. (2005) Quorum sensing and quorum-quenching enzymes, *J. Microbiol.* 43, 101–9.
- Rice, S. A., McDougald, D., Kumar, N., and Kjelleberg, S. (2005) The use of quorum-sensing blockers as therapeutic agents for the control of biofilm-associated infections, *Curr. Opin. Invest. Drugs* 6, 178–84.
- Suga, H., and Smith, K. M. (2003) Molecular mechanisms of bacterial quorum sensing as a new drug target, *Curr. Opin. Chem. Biol.* 7, 586–91.
- Lin, Y. H., Xu, J. L., Hu, J., Wang, L. H., Ong, S. L., Leadbetter, J. R., and Zhang, L. H. (2003) Acyl-homoserine lactone acylase from *Ralstonia* strain XJ12B represents a novel and potent class of quorum-quenching enzymes, *Mol. Microbiol.* 47, 849–60.
- Huang, J. J., Han, J. I., Zhang, L. H., and Leadbetter, J. R. (2003) Utilization of acyl-homoserine lactone quorum signals for growth by a soil pseudomonad and *Pseudomonas aeruginosa* PAO1, *Appl. Environ. Microbiol.* 69, 5941–9.
- Park, S. Y., Kang, H. O., Jang, H. S., Lee, J. K., Koo, B. T., and Yum, D. Y. (2005) Identification of extracellular N-acylhomoserine lactone acylase from a *Streptomyces* sp. and its application to quorum quenching, *Appl. Environ. Microbiol.* 71, 2632–41.
- Yang, F., Wang, L. H., Wang, J., Dong, Y. H., Hu, J. Y., and Zhang, L. H. (2005) Quorum quenching enzyme activity is widely conserved in the sera of mammalian species, *FEBS Lett.* 579, 3713–7.
- Draganov, D. I., Teiber, J. F., Speelman, A., Osawa, Y., Sunahara, R., and La Du, B. N. (2005) Human paraoxonases (PON1, PON2, and PON3) are lactonases with overlapping and distinct substrate specificities, *J. Lipid Res.* 46, 1239–47.
- Dong, Y. H., Gusti, A. R., Zhang, Q., Xu, J. L., and Zhang, L. H. (2002) Identification of quorum-quenching N-acyl homoserine lactonases from *Bacillus* species, *Appl. Environ. Microbiol.* 68, 1754–9.
- Lee, S. J., Park, S. Y., Lee, J. J., Yum, D. Y., Koo, B. T., and Lee, J. K. (2002) Genes encoding the N-acyl homoserine lactone-degrading enzyme are widespread in many subspecies of *Bacillus thuringiensis*, *Appl. Environ. Microbiol.* 68, 3919–24.
- Dong, Y. H., Xu, J. L., Li, X. Z., and Zhang, L. H. (2000) AiiA, an enzyme that inactivates the acylhomoserine lactone quorum-sensing signal and attenuates the virulence of *Erwinia carotovora*, *Proc. Natl. Acad. Sci. U.S.A.* 97, 3526–31.
- Reimann, C., Ginot, N., Michel, L., Keel, C., Michaux, P., Krishnapillai, V., Zala, M., Heurlier, K., Triandafillu, K., Harms, H., Defago, G., and Haas, D. (2002) Genetically programmed autoinducer destruction reduces virulence gene expression and swarming motility in *Pseudomonas aeruginosa* PAO1, *Microbiology* 148, 923–32.

16. Ulrich, R. L. (2004) Quorum quenching: Enzymatic disruption of N-acylhomoserine lactone-mediated bacterial communication in *Burkholderia thailandensis*, *Appl. Environ. Microbiol.* **70**, 6173–80.
17. Wopperer, J., Cardona, S. T., Huber, B., Jacobi, C. A., Valvano, M. A., and Eberl, L. (2006) A quorum-quenching approach to investigate the conservation of quorum-sensing-regulated functions within the *Burkholderia cepacia* complex, *Appl. Environ. Microbiol.* **72**, 1579–87.
18. Liu, D., Lepore, B. W., Petsko, G. A., Thomas, P. W., Stone, E. M., Fast, W., and Ringe, D. (2005) Three-dimensional structure of the quorum-quenching N-acyl homoserine lactone hydrolase from *Bacillus thuringiensis*, *Proc. Natl. Acad. Sci. U.S.A.* **102**, 11882–7.
19. Thomas, P. W., Stone, E. M., Costello, A. L., Tierney, D. L., and Fast, W. (2005) The quorum-quenching lactonase from *Bacillus thuringiensis* is a metalloprotein, *Biochemistry* **44**, 7559–69.
20. Kim, M. H., Choi, W. C., Kang, H. O., Lee, J. S., Kang, B. S., Kim, K. J., Derewenda, Z. S., Oh, T. K., Lee, C. H., and Lee, J. K. (2005) The molecular structure and catalytic mechanism of a quorum-quenching N-acyl-L-homoserine lactone hydrolase, *Proc. Natl. Acad. Sci. U.S.A.* **102**, 17606–11.
21. Wang, L. H., Weng, L. X., Dong, Y. H., and Zhang, L. H. (2004) Specificity and enzyme kinetics of the quorum-quenching N-acyl homoserine lactone lactonase (AHL-lactonase), *J. Biol. Chem.* **279**, 13645–51.
22. Garrity, J. D., Bennett, B., and Crowder, M. W. (2005) Direct evidence that the reaction intermediate of metallo- β -lactamase L1 is metal bound, *Biochemistry* **44**, 1078–87.
23. Glasoe, P. K., and Long, F. A. (1960) Use of glass electrodes to measure acidities in deuterium oxide, *J. Phys. Chem.* **64**, 188–9.
24. Chhabra, S. R., Harty, C., Hooi, D. S., Daykin, M., Williams, P., Telford, G., Pritchard, D. I., and Bycroft, B. W. (2003) Synthetic analogues of the bacterial signal (quorum sensing) molecule N-(3-oxododecanoyl)-L-homoserine lactone as immune modulators, *J. Med. Chem.* **46**, 97–104.
25. Periyannan, G. R., Costello, A. L., Tierney, D. L., Yang, K. W., Bennett, B., and Crowder, M. W. (2006) Sequential binding of cobalt(II) to metallo- β -lactamase CcrA, *Biochemistry* **45**, 1313–20.
26. Ankudinov, A. L., Ravel, B., Rehr, J. J., and Conradson, S. D. (1998) Real space multiple scattering calculation and interpretation of XANES, *Phys. Rev. B* **58**, 7565–76.
27. Risley, J. M., and Van Etten, R. L. (1989) Mechanistic studies utilizing oxygen-18 analyzed by carbon-13 and nitrogen-15 nuclear magnetic resonance spectroscopy, *Methods Enzymol.* **177**, 376–89.
28. Llewellyn, D. R., and O'Connor, C. (1964) Tracer studies of carboxylic acids. Part I. Acetic and pivalic acid, *J. Chem. Soc.*, 545–9.
29. Risley, J. M., and Van Etten, R. L. (1981) ^{18}O -isotope effect in ^{13}C nuclear magnetic resonance spectroscopy. 4. Oxygen exchange of $[\text{1-}^{13}\text{C}, ^{18}\text{O}_2]\text{acetic acid}$ in dilute acid, *J. Am. Chem. Soc.* **103**, 4389–92.
30. Rehm, T., Huber, R., and Holak, T. A. (2002) Application of NMR in structural proteomics: Screening for proteins amenable to structural analysis, *Structure* **10**, 1613–8.
31. Wüthrich, K. (1986) *NMR of proteins and nucleic acids*, Wiley, New York.
32. Wang, Z., and Benkovic, S. J. (1998) Purification, characterization, and kinetic studies of a soluble *Bacteroides fragilis* metallo- β -lactamase that provides multiple antibiotic resistance, *J. Biol. Chem.* **273**, 22402–8.
33. Neurath, H. (1970) *The Proteins: Composition, structure, and function*, 2nd ed., Vol. 5, Academic Press, New York.
34. Bond, M. D., Holmquist, B., and Vallee, B. L. (1986) Thioamide substrate probes of metal-substrate interactions in carboxypeptidase A catalysis, *J. Inorg. Biochem.* **28**, 97–105.
35. Sillén, L. G., and Martell, A. E. (1971) *Stability constants of metal-ion complexes. Supplement*, Royal Chemical Society, London.
36. Sillén, L. G., Martell, A. E., and Bjerrum, J. (1964) *Stability constants of metal-ion complexes*, 2nd ed., Royal Chemical Society, London.
37. Kaiser, E. T., and Kezdy, F. J. (1976) Hydrolysis of Cyclic Esters, *Prog. Bioorg. Chem.* **4**, 239–67.
38. Huisgen, R., and Ott, H. (1959) Die konfiguration der carbonestergruppe und die sondereigenschaften der lactone, *Tetrahedron* **6**, 253–67.
39. Johnston, N. J., Mukhtar, T. A., and Wright, G. D. (2002) Streptogramin antibiotics: Mode of action and resistance, *Curr. Drug Targets* **3**, 335–44.
40. Tobias, P. S., and Kezdy, F. J. (1969) The alkaline hydrolysis of 5-nitrocoumaranone. A method for determining the intermediacy of carbanions in the hydrolysis of esters with labile α protons, *J. Am. Chem. Soc.* **91**, 5171–3.
41. Bruice, T. C., and Holmquist, B. (1969) The carbanion (E1cB) mechanism of ester hydrolysis. I. hydrolysis of malonate esters, *J. Am. Chem. Soc.* **91**, 2993–3002.
42. Moore, J. A., and Schwab, J. M. (1991) Unprecedented observation of lactone hydrolysis by the AAI2 mechanism, *Tetrahedron Lett.* **32**, 2331–4.
43. Dong, Y. H., Wang, L. H., Xu, J. L., Zhang, H. B., Zhang, X. F., and Zhang, L. H. (2001) Quenching quorum-sensing-dependent bacterial infection by an N-acyl homoserine lactonase, *Nature* **411**, 813–7.
44. Dragani, B., Cocco, R., Ridderstrom, M., Stenberg, G., Mannervik, B., and Aceto, A. (1999) Unfolding and refolding of human glyoxalase II and its single-tryptophan mutants, *J. Mol. Biol.* **291**, 481–90.
45. Periyannan, G., Shaw, P. J., Sigdel, T., and Crowder, M. W. (2004) In vivo folding of recombinant metallo- β -lactamase L1 requires the presence of Zn(II), *Protein Sci.* **13**, 2236–43.
46. Daiyasu, H., Osaka, K., Ishino, Y., and Toh, H. (2001) Expansion of the zinc metallo-hydrolase family of the β -lactamase fold, *FEBS Lett.* **503**, 1–6.
47. Hagelueken, G., Adams, T. M., Wiehlmann, L., Widow, U., Kolmar, H., Tummler, B., Heinz, D. W., and Schubert, W. D. (2006) The crystal structure of SdsA1, an alkylsulfatase from *Pseudomonas aeruginosa*, defines a third class of sulfatases, *Proc. Natl. Acad. Sci. U.S.A.* **103**, 7631–6.
48. Aubert, S. D., Li, Y., and Raushel, F. M. (2004) Mechanism for the hydrolysis of organophosphates by the bacterial phosphotriesterase, *Biochemistry* **43**, 5707–15.
49. Murphy, B. P., and Pratt, R. F. (1989) A thiono- β -lactam substrate for the β -lactamase II of *Bacillus cereus*. Evidence for direct interaction between the essential metal ion and substrate, *Biochem. J.* **258**, 765–8.
50. Loppreore, C., and Byers, L. D. (1998) The urease-catalyzed hydrolysis of thiourea and thioacetamide, *Arch. Biochem. Biophys.* **349**, 299–303.
51. Bienvenue, D. L., Gilner, D., and Holz, R. C. (2002) Hydrolysis of thiono-peptides by the aminopeptidase from *Aeromonas proteolytica*: Insight into substrate binding, *Biochemistry* **41**, 3712–9.
52. Siegel, H., and McCormic, D. B. (1970) On the discriminating behavior of metal ions and ligands with regard to their biological significance, *Acc. Chem. Res.* **3**, 201–8.
53. Spencer, J., Clarke, A. R., and Walsh, T. R. (2001) Novel mechanism of hydrolysis of therapeutic β -lactams by *Stenotrophomonas maltophilia* L1 metallo- β -lactamase, *J. Biol. Chem.* **276**, 33638–44.

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